Sarothrin from Alkanna orientalis Is an Antimicrobial Agent and Efflux Pump Inhibitor

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Abstract:

An Alkanna orientalis leaf and flower extract inhibited the growth of Staphylococcus aureus, a pathogen that causes an estimated 478000 hospitalizations in the US annually. Bioassay-guided fractionation of A. orientalis resulted in isolation of the flavonoid sarothrin (5,7,4′-trihydroxy-3,6,8-trimethoxyflavone), which inhibited the growth of Mycobacterium smegmatis (MIC 75 μM) and S. aureus (MIC > 800 μM), and possessed efflux pump inhibitory activity. This is the first report of antimicrobial or efflux pump inhibitory activity of sarothrin, and of its presence in A. orientalis. Our findings suggest that the effectiveness of A. orientalis extracts is due to a combination of multiple constituents, including sarothrin.

Keywords: Alkanna orientalis | Boraginaceae | antimicrobial | efflux inhibition

Article:

Bacterial infections have an estimated $20 billion burden on the US health care system [1]. Botanicals have been suggested as an under-utilized source of antimicrobial agents [2, 3]. With this project, our goals were to evaluate the antimicrobial activity of the plant Alkanna orientalis (L.) Boiss (Boraginaceae) against Staphylococcus aureus and Mycobacterium smegmatis, and to identify compounds that play a role in this activity. A. orientalis was chosen for this study based on antimicrobial activity observed for the crude extract by our laboratory and others [4, 5], and on the ethnobotanical literature. This plant genus was traditionally employed as a treatment for digestive problems [4] and for wound healing.

Bioassay-guided fractionation of Alkanna orientalis resulted in the isolation of the flavonoid sarothrin (5,7,4′-trihydroxy-3,6,8-trimethoxyflavone) (Fig. 1). Sarothrin is present in other botanicals, including Encelia densifolia (Asteraceae) [6], Ononis rotundifolia (Fabaceae), and Gardenia obtusifolia (Rubiaceae) [7]. However, this is the first report of sarothrin in A. orientalis or any member of the Boraginaceae family.

Sarothrin was observed to inhibit M. smegmatis (MIC 75 μM) and weakly inhibited S. aureus growth [MIC > 800 μM, Table 1, 50% inhibition of growth at 38 μg/mL (100 μM), Fig. 3S]. However, the crude A. orientalis leaf and flower extract, which contained only 1.63 ± 0.13% sarothrin, had very similar activity to that of sarothrin alone (Fig. 3S). Furthermore, comparisons were made of sarothrin concentrations in various A. orientalis plant parts (Table 2). The highest levels were extracted from leaves and flowers, while very lowlevels were present in roots and seeds (Table 2). Nonetheless, similar antimicrobial activity (30 to 60% inhibition) was observed from extracts of various plant parts (Fig. 3S). Collectively, these findings suggest that additional constituents besides sarothrin are likely to play a role in the antimicrobial activity of A. orientalis.

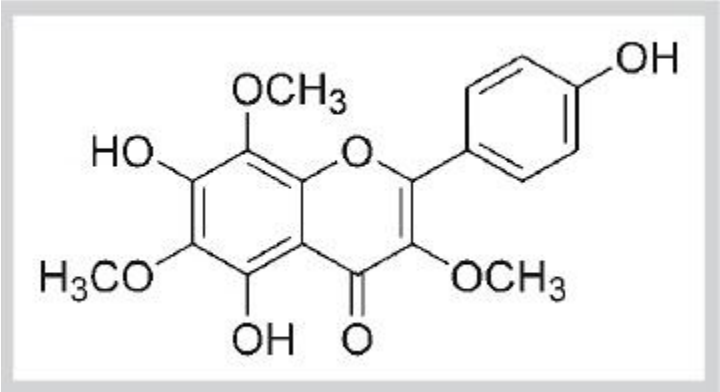

Fig. 1 Structure of sarothrin (1) isolated from Alkanna orientalis as a result of bioactivity-directed fractionation evaluating antimicrobial activity against Staphylococcus aureus.

Table 1. MIC (concentration required to completely inhibit bacterial growth) measured for purified sarothrin against two pathogenic bacteria.

Organism	**MIC sarothrin (μM)**	**MIC ciprofloxacin (μM)**
Staphylococcus aureus NCTC 8325‑4	> 800	1.5
Mycobacterium smegmatis ATCC 607	75	6

Table 2. Quantity of the bioactive flavonoid sarothrin in extracts prepared from various plant parts of Alkanna orientalis.

Plant part	**Sarothrin concentration (ppm)[a] ± SD**
Root[b]	0.51 ± 0.40
Seed	0.37 ± 0.12
Leaf	52.9 ± 1.4
Flower + leaf	160 ± 13

[a]Quantities are reported as mg sarothrin/kg plant material based on LC‑MS analysis of extracts prepared from the relevant plant parts. Standard deviations are for triplicate analyses of the same extract. Error bars represent ± the standard deviation of each extract concentration based on linear regression analysis of a 9 point calibration curve of peak area versus concentration with slope (m) = 0.9787 ± 0.0021, intercept (b) = 6.346 ± 0.018, and R2 = 0.9967;

[b] The concentration reported for the root extract is approximate only, as the concentration in the extract at the dilution analyzed was below the lower limit of detection for the method. Concentrations for the seed, leaf, and flower + leaf extract fell within the linear range of the calibration curve.

Efflux pump inhibition in combination with antibiotics has been proposed as a potential therapeutic strategy against bacterial infections [3]. Reports indicating efflux pump inhibitory activity of flavonoids [8–11] led us to investigate the efflux pump inhibitory activity of sarothrin. A fluorescence-based assay was utilized, which relies on the efflux of ethidium bromide driven by the S. aureus efflux pump NorA [12]. As is apparent from the data in Fig. 2, sarothrin blocked ethidium bromide efflux (data overlaid with the positive control CCCP). These findings suggest that sarothrin possesses efflux pump inhibitory activity. This could be relevant to the overall effectiveness of A. orientalis extracts against bacteria; while sarothrin is only a weak antimicrobial agent alone, it could increase the activity of other antimicrobial compounds in the extracts by blocking bacterial efflux pumps.

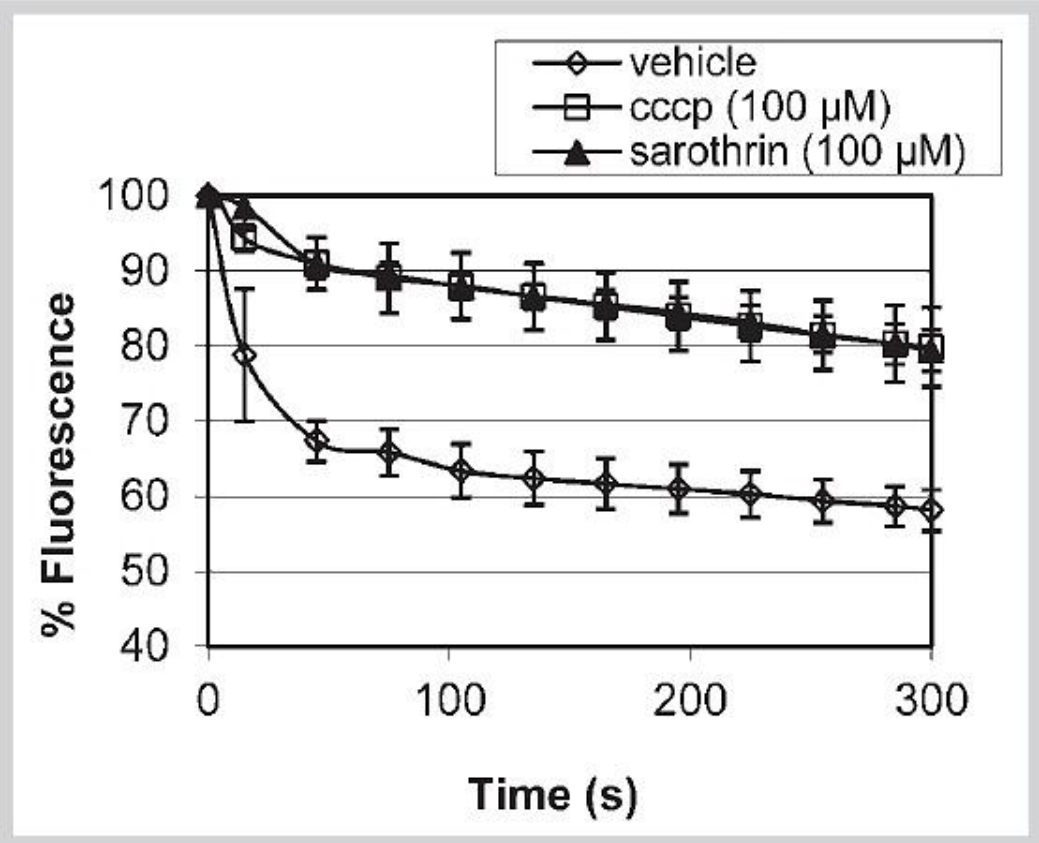

Fig. 2 Percent fluorescence over time for S. aureus (NCTC 8325‑4) loaded with ethidium bromide and treated with purified sarothrin. The known efflux pump inhibitor CCCP (carbonyl cyanide m-chlorophenylhydrazone) served as a positive control. Vehicle consisted of 10% DMSO in Müller Hinton broth. Triplicate measurements were made for separate aliquots of solution with different S. aureus pellets, and data points represent the average of these three

measurement. Error bars are ± standard error. Fluorescence measurements were made using λex = 530 nm, λem = 600 nm.

Materials and Methods

Staphylococcus aureus (NCTC 8325-4) [13] and Mycobacterium smegmatis (ATCC 607) were employed. Müller Hinton broth, carbonyl cyanide m-chlorophenylhydrazone (CCCP), berberine, and ciprofloxacinwere purchased from Sigma-Aldrich, all with purity > 98%.

Alkanna orientalis was cultivated at Horizon Herbs and identified by Richard Cech. A voucher was deposited in the University of North Carolina Herbarium (NCU 592736). Dried, powdered samples from A. orientalis leaves (2.0 g), roots (2.0 g), leaves + flowers (2060 g), or seeds (10.5 g)were extracted in methanol (1 : 12.5, w/v). Extracts were stirred for 24 hr, filtered and rotary evaporated. The residue was separated with liquid/liquid partitioning, as described elsewhere [14]. Final yields of the organic fraction were 17.4mg, 7.3mg, 20.3 g, and 1.5mg, respectively, for the leaf, root, flower + leaf, and seed extracts. The flower + leaf extract was fractionated over silica gel with a hexane: chloroform:methanol gradient as described [15]. The most active fraction (strongest inhibition of S. aureus) was separated over silica gel utilizing hexane: ethyl acetate as the gradient [15]. Yellow crystals (52.0mg, 92% pure) (sarothrin) precipitated and were purified using reversed-phase preparative HPLC with a YMC ODS‑A column (5 μm, 120 Å; 250 × 20 mm; Waters) with a CH3CN:H2O gradient. Sarothrin (Fig. 1) eluted at 13.5min (7.03mg, 97% purity, 0.00034% yield).

Sarothrin (5,7,4′-trihydroxy-3,6,8-trimethoxyflavone) (1): yellow solid, HRESIMS 361.09100 [M + H]+ (calcd. for C18H17O8, 361.09 180); 1H NMR (500 MHz acetone-d6) (Fig. 1S) and 13C NMR (125 MHz, acetone-d6) (Fig. 2S) agreed with literature values (Table 1S) [16]. The HRMS and NMR instruments employed were an LTQ-Orbitrap (Thermo) and JEOL ECA-500, respectively. Sarothrin quantitation was performed using a triple quadrupole mass spectrometer (TSQ Access; Thermo) with positive ion electrospray coupled to an HP1200 HPLC (Agilent) with a C-18 Prevail column. An acetonitrile (1% formic acid) :water (1% formic acid) gradient was employed at 0.3mL/min.

Mycobacterium smegmatis was grown in Middlebrook 7H9 medium, and MIC values measured after 3 days incubation were as described previously [17]. Staphylococcus aureus was grown in Müller Hinton broth, MICs measured using CLSI standard methods [18], and efflux pump inhibitory activity evaluated, as described previously [15,19].

Supporting information

NMR data for sarothrin and comparison of S. aureus growth inhibition by various A. orientalis extracts are available as Supporting Information.

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Conflict of Interest

None

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