HPLC Method Validation: A Global Application for the Analysis of Amoxicillin

Senior Project

In partial fulfillment of the requirements for
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University of North Carolina at Pembroke

By

Ethan Williamson
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Ethan Williamson
Honors College Scholar

Meredith Storms, Ph.D.
Faculty Mentor

Teagan Decker, Ph.D.
Senior Project Coordinator

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Several factors can determine the purity and efficacy of certain pharmaceutical compounds. These environmental factors play a huge role in how drug manufacturers store and handle compounds. Several countries that are without resources such as dependable electricity and clean water, also suffer from not receiving proper healthcare, which includes not receiving correct medication. **The main goal of this research is to validate the HPLC method which will be used to analyze the purity of amoxicillin capsules collected from selected African countries to show the importance of safe and effective pharmaceutical products.** Before the samples can be analyzed, it is necessary to meet several criteria for a validated HPLC method. **Thus, the aim of this research is to demonstrate the linearity of the HPLC method as well as to show that the method meets the requirements for tailing factor and column efficiency.**
Background

Amoxicillin is in the beta lactam family that also belongs to the Penicillins, which can be used to treat an overabundance of infections including: pneumonia, ear infections, bronchitis, and tonsillitis. Amoxicillin became available when it was introduced to the United Kingdom in 1972. Amoxicillin is dispensed in a variety of forms from tablets to liquid drops, to be taken orally. These various dispensing techniques makes it difficult to manage the integrity of the drug. Amoxicillin is listed on the World Health Organization’s “List of Essential Medicines” as one of the most vital medications needed in a basic health system. Amoxicillin is one of the most frequently prescribed antibiotic used to treat infections in patients who are commonly children. However, Amoxicillin that is distributed in third world countries, like Africa, have different standards for handling and storage compared to the United States which may impact the efficacy of the drug. This is due to environmental factors, including temperature, humidity, storage containers/packaging, light, and humidity which degrade the purity of the medication. These medications are simply not as effective as pharmaceutical drugs in the United States.

This outbreak has caught the attention of researcher’s everywhere which led University of Notre Dame and the Moi Teaching and Referral Hospital to create a research trial that would test the purity of these samples taken from street pharmacies in West Kenyan Communities by covert shoppers. The samples are collected and sent to the Moi Teaching and Referral Hospital, where they are...
Williamson cataloged and then sent to University of Notre Dame. The samples are then boxed and sent to the participating colleges that will be conducting the testing of the samples. However, this research is considered a blind study, which means that the participating institution does not know if they are testing a sample collected from Africa or a placebo created by the oversight institution. This allows for the integrity of the research to be carried out and for no discrimination to occur in both the researcher or the participating institution. There are certain parameters met to fulfill the requirements to conduct this research.

**Methodology**

To conduct this research, a reversed-phase high-performance liquid chromatography (HPLC) method with UV detection, the subsequent components of these drugs are able to be compared to the original compound to differentiate whether or not they contain the same properties. The HPLC machine must be calibrated and the accuracy insured to be operating within the right guidelines. Precision, linearity, and accuracy are the three things required to test and fully satisfy the protocol established by University of Notre Dame of the Amoxicillin provided. The machine is set to a wavelength of 230 nm, a monosodium phosphate buffer is used at a 0.5 mL/min injection rate. These guidelines are constant for every injection. To test these parameters, pharmaceutical grade Amoxicillin is used and certain dilutions are created to test the linearity of the compound. These samples are then injected (20 µL injection volume) into the chromatography column in the HPLC machine and the machine generates a graph that lists the peak area, peak height, theoretical plate, tailing factor, and k prime data. The data illustrates how the compound breaks down over a 15 minute time period which also shows how the pure Amoxicillin breaks
Williamson
down compared to counterfeit samples. This same procedure is completed to
calculate the precision part of the data. Compared to linearity, the precision aspect
measures the retention time, the time it takes for Amoxicillin to elute from the
column. Instead of using different concentration of Amoxicillin, the same
concentration was used repeatedly. Also, an optional test to validate is the Limit of
Detection (LOD) and Lower Limits of Quantification (LLOQ) which is defined as the
lowest concentration at which 95% of positive samples are detected. Simply, this
shows the smallest concentration of Amoxicillin that is detectable in the HPLC
machine. These above mentioned test areas have specific guidelines concerning the
acceptance or rejection of data, which points the researchers in the “right” direction
of the calibration of their machines. Without these guidelines, it would be
impossible to fully accept any data collected from other machines since each would
be set up so differently.

Results and Discussion

According to Dr. Liberman, designer of the analytical method used in this research,
she recommended to “…prepare and run at least five calibration standards over the
concentration range of 5% to 200%.” After reviewing the data as a whole, a graph
Williamson was created. Each single data point on the calibration curve represents triplicate injections of various amoxicillin concentrations. Based on the method requirements, the acceptable $R^2$ point is to be $\geq 0.98$. As noted on the calibration curve, the method resulted in a $R^2$ value of 0.99 which is creditable.

### Quality Control Chart

<table>
<thead>
<tr>
<th>Date</th>
<th>Theoretical Plates</th>
<th>Tailing Factor</th>
<th>$k'$</th>
</tr>
</thead>
<tbody>
<tr>
<td>11-Feb</td>
<td>3808.634</td>
<td>1.133</td>
<td>2.024</td>
</tr>
<tr>
<td>18-Feb</td>
<td>3111.859</td>
<td>1.185</td>
<td>1.523</td>
</tr>
<tr>
<td>18-Feb</td>
<td>3148.853</td>
<td>1.198</td>
<td>1.522</td>
</tr>
<tr>
<td>18-Feb</td>
<td>3028.968</td>
<td>1.240</td>
<td>1.554</td>
</tr>
<tr>
<td>19-Mar</td>
<td>2724.039</td>
<td>1.009</td>
<td>1.041</td>
</tr>
</tbody>
</table>

To ensure the validation of the method, an external standard is used to ascertain various quality control parameters.

- The number of **theoretical plates** must be $> 1700$.
- The **tailing factor** must be $< 2.5$.
- The **retention factor ($k'$)** must be between 1.1-2.2.

Based on the data presented in the Quality Control Chart, all three criteria are met with this HPLC method.

### Precision Table

<table>
<thead>
<tr>
<th>Run</th>
<th>Peak Area</th>
<th>Retention Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2848397</td>
<td>6.796</td>
</tr>
<tr>
<td>2</td>
<td>2913860</td>
<td>6.552</td>
</tr>
<tr>
<td>3</td>
<td>2927961</td>
<td>6.608</td>
</tr>
<tr>
<td>4</td>
<td>2967067</td>
<td>6.760</td>
</tr>
<tr>
<td>5</td>
<td>3005648</td>
<td>6.695</td>
</tr>
<tr>
<td>6</td>
<td>2895903</td>
<td>6.768</td>
</tr>
</tbody>
</table>

**Mean:** 2926472.667  
**Standard Deviation:** 54969.523
The Relative Standard Deviation (RSD) for the integrated intensities of 6 consecutive injections of the known standard should be below 2%. The RSD found from our study was 1.88%, so the parameters were met. The peak area is the area from the chromatogram. The retention time is the time it takes for Amoxicillin to elute from the column. The range of the retention time is less than 0.5 minutes, which is another parameter that was met.

<table>
<thead>
<tr>
<th>Concentration</th>
<th>Run 1 (peak area)</th>
<th>Run 2 (peak area)</th>
<th>Run 3 (peak area)</th>
<th>Average Peak Area</th>
<th>Std Dev of Peak Area</th>
<th>%RSD</th>
</tr>
</thead>
<tbody>
<tr>
<td>6 µg/mL</td>
<td>1411810</td>
<td>1417620</td>
<td>1439726</td>
<td>1423052</td>
<td>14729.417 23</td>
<td>1.0350582 57</td>
</tr>
<tr>
<td>10 µg/mL</td>
<td>2294849</td>
<td>2366629</td>
<td>2378886</td>
<td>2346788</td>
<td>45396.071 45</td>
<td>1.9343916 64</td>
</tr>
<tr>
<td>10 µg/mL</td>
<td>2411442</td>
<td>2405287</td>
<td>2424846</td>
<td>2413858.3 33</td>
<td>10000.880 98</td>
<td>0.4143110 157</td>
</tr>
<tr>
<td>15 µg/mL</td>
<td>3348757</td>
<td>3396954</td>
<td>3320738</td>
<td>3355483</td>
<td>38550.602 73</td>
<td>1.1488838 64</td>
</tr>
<tr>
<td>20 µg/mL</td>
<td>4904042</td>
<td>4902144</td>
<td>4933249</td>
<td>4913145</td>
<td>17436.419 16</td>
<td>0.3548932 335</td>
</tr>
</tbody>
</table>
The LOD/LLOQ was determined by using varying concentrations of Amoxicillin each at a smaller concentration until it was undetectable by the HPLC machine. The %RSD will also be measured and has to be under 2%. Since each value was under the mentioned value, we were able to accept each one and check off the LOD/LLOQ portion of the test.

**Conclusion**

The purpose of our research is to validate the HPLC machine which will be used to analyze Amoxicillin samples purchased in African countries. At this point, our research demonstrates that the linearity of the data (0.999 $R^2$), quality and efficacy of our standards, parameters for precision (1.88% RSD), and the range for retention time (0.244 min) has been satisfied. We also determined the LOD to be 0.6 $\mu$g/mL with the LOQ equivalent to 10 $\mu$g/mL. Our results from the precision
Williamson study met the specific parameters allowing us to continue with our research. After final validation and approval, our hopes are to begin testing the samples which will then be sent to the Medical Regulatory Authority (MRA) and the WHO Medical Rapid Alert System (MRA).

References

Williamson


