Landiolol Hydrochloride Ameliorates Acute Lung Injury In A Rat Model Of Early Sepsis Through The Suppression Of Elevated Levels Of Pulmonary Endothelin-1

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Abstract

Among the dysfunctions and pathologies associated with sepsis, the underlying molecular mechanisms of sepsis-induced acute lung injury (ALI) are poorly understood. Endothelin (ET)-1, a potent vasoconstrictor and pro-inflammatory peptide, is known to be involved in the pathogenesis of ALI in a rat model of sepsis. Here, we investigated whether landiolol hydrochloride, an ultra-short-acting β-blocker, plays a crucial role in ameliorating and attenuating LPS-induced ALI through modulation of the ET-1 system. Male Wistar rats at 8 weeks of age were ad-ministered with either saline or lipopolysaccharide (LPS) for three hours (3 h) and some of the LPS-administered rats were continuously treated with landiolol for 3 h. ALI was induced by LPS, including levels of both circulatory and pulmonary TNF-α and IL-6 but \([\text{PaO}_2]\) was significantly decreased. LPS also induced a significant increase in levels of pulmonary ET-1 and ET-A receptor, but levels of ET-B receptor, which has vasodilating effects, were remarkably diminished. Further, LPS administration upregulated the pulmonary expression of HIF-1α. Finally, the treatment of LPS-administered rats with landiolol for 3 h ameliorated and prevented ALI, normalized the altered levels of pulmonary ET-1 and ET-A receptors. Landiolol also induced significant down-regulation of ET-B receptor in lung tissues in the early hours (phase) of sepsis. However, Landiolol treatment had no effect on the up-regulated inflammatory mediators (TNF-α, IL-6) in both plasma and lung tissues during sepsis, and expression of pulmonary HIF-1α also remained unchanged after landiolol treatment. Collectively, these data led us to conclude that landiolol may ameliorate sepsis-induced ALI via the pulmonary ET system.

1. Introduction

Sepsis is a critical life-threatening condition with a definition that encompasses pathologic infection and physiological changes that are collectively known as systemic inflammatory response syndrome (SIRS) [1]. The mortality from sepsis and the associated complications is very high and is estimated to be about 30% [2], making it the second leading cause of death among patients admitted at non-coronary intensive care units [3]. To date, the pathogenesis of sepsis and its progression to multiple organ dysfunction syndrome (MODS) and the associated septic shock are poorly understood and thus have been the subject of investigations in the last several decades.

Acute lung injury (ALI) is a well-known and frequent complication of sepsis, and causes a significant number of sudden deaths (mortality) and morbidity [4–6]. For this reason, patients with ALI/ARDS account for a significant proportion of the intensive care unit (ICU) case load. ALI and acute respiratory distress syndrome (ARDS), the severe form of ALI, are characterized by an acute onset of severe hypoxia pulmonary infiltration [7], pulmonary hypertension, edema and deteriorated gas exchange [8]. Indeed, sepsis is a potential risk factor for ALI and ARDS [5].

To date, the molecular mechanisms underlying the pathogenesis of sepsis-induced ALI are poorly defined and understood. We do know that endothelin (ET)-1, a potent endogenous vasoconstrictor and pro-
inflammatory peptide [9,10], is actively involved in the pathogenesis of sepsis and the sepsis-associated organ dysfunction, as well as the associated complications [11], as evidenced by data from recent studies. Specifically, plasma levels of ET-1 are elevated in various sepsis animal models, including septic patients [12–15] and a clear correlation has been observed between ET plasma levels and morbidity/mortality in septic patients. These observations implicate ET in the pathogenesis of septic shock in human [16,17]. Further, ET-1 likely has a direct role in the development and subsequent severity of ALI by increasing the pressure of pulmonary microvasculature during the first phase or hours of sepsis [18,19]. Interestingly, endothelin blocker, namely tezosentan, ameliorates pulmonary hypertension, lung edema, cardiac dysfunction, and arterial hypoxemia in an ovine model of endotoxin-induced lung injury [20]. In contrast, intravenous infusion of ET-1 causes an elevation in pulmonary artery pressure and edema [21], implying a direct role of ET-1 in the pathogenesis of lung injury.

Landiolol hydrochloride, an ultra-short-acting and highly cardio-sele-
tive beta-1 blocker, with a half-life of 4 min, has been used in treating several acute medical disorders, including arrhythmias, during heart surgery [22], acute myocardial infarction [23], acute decompensated heart failure [24], and refractory electrical storm [25]. Ultra-short-acting beta-blockers, such as landiolol, can influence heart rate but exert minimal effect on cardiac function. Besides, the potential effects of landiolol on rhythm control, landiolol also plays a protective role against ALI in a rat model of lipopolysaccharide (LPS)-induced systemic inflammation, which is associated with a reduction in high mobility group box 1 (HMGB-1) [26]. Very recently, we have demonstrated that landiolol is effective in improving acute liver injury of sepsis through the modulation of TNF (tumor necrosis factor-alpha)-α [27]. In addition, we have also shown that ET-1 is highly upregulated in kidney and heart tissues in LPS-administered rats [28,29] and landiolol has inhibitory effects on the upregulated levels of ET-1 in both of these tissues. However, it is not known whether landiolol will equally exert protective effects in lung tissues during sepsis through the alteration of the ET-1 system. Indeed, in our previous study we have already reported elevated ET-1 levels in lung tissues during sepsis [30].

In the present study, we investigated whether landiolol hydrochloro-
ide, an ultra-short-acting beta-blocker, can play an important role in attenuating LPS-induced ALI through the modulation of pulmonary ET-1.

2. Materials and methods

Male Wistar rats (200–250 g, 8 weeks old) were used in all experiments in the current study. Sepsis was induced by the intra-peritoneal (IP) administration of bacterial LPS from Escherichia coli O55: B5 (15 mg/kg), dissolved in sterile saline, as described previously [31–33]. A dose 15 mg/kg of LPS has been shown to induce morphological injuries in lung [34] as shown in our dose response study.

The total number of rats used in all the experiment was 45, which was randomized into three groups, namely: Group 1 (control, n = 15), group 2 (LPS, n = 15) and group 3 (LPS + landiolol hydrochloride, n = 15). For group 1, sterile saline (2 ml/body) was administered at time 0 h and then the rats were killed after 3 h (control group). For group 2, LPS at a dose of 15 mg/kg was administered at 0 h, and then the rats were killed after 3 h (sepsis group). In group 3, landiolol hydrochloride was administered intravenously (100 μg/kg/min) for 15 min non-stop before LPS administration (landiolol treated sepsis group). The dose for landiolol was found to be the minimal dose for normalizing the LPS-induced hyperdynamic state in the acute (early) phase (hours) of sepsis, as reported in our past study [28].

Nembutal (sodium pentobarbital, IP, 80 mg/kg body weight, rou-
tinely used in our lab) was used to kill all the rats at 3 h, at the end of the experimental protocol. Blood gas analysis was also performed in the current study. The blood samples were collected from a polypropylene tube catheter inserted into the left carotid artery for blood gas analysis, and then lung tissues were carefully harvested, snap-frozen in liquid nitrogen, and stored at −80 °C. All animals received proper care and the experimental procedures were approved by the Animal Care and Use Committee of University of Tsukuba prior to the study.

2.1. Measurements of hemodynamic parameters

Hemodynamic parameters were measured using a previously optim-
ized methodology from our lab [27–35]. Briefly, rats were anesthe-
tized with isoflurane inhalation (1.5%, 1 l/min) and a microtip pressure transducer catheter (SPC-320, Millar Instruments, Houston, TX, USA) was inserted into the left carotid artery at the end of the experimental protocol. Arterial blood pressure and heart rate (HR) were monitored with a pressure transducer (model SCK-590, Gould, Ohio, USA) and recorded with the use of a polygraph system (amplifier, AP-601G, Nihon Kohden, Tokyo, Japan; Tachometer, AT-601G, Nihon Kohden; and thermal-pen recorder, WT-687G, Nihon Kohden).

2.2. Echocardiography

Echocardiography was performed using a Vevo 2100 high-frequency ultrasound system (VisualSonics, Inc., Ontario, Canada), which includes an integrated rail system for consistent positioning of the ultrasound probe [36] at the end of the protocol. The fur from the chest was shaved with an electrical clipper and a gel. The animals were then connected to an electrocardiogram (ECG). An optimal parasternal long axis (LAX) cine loop (i.e. visualization of both the mitral and aortic valves, and maximum distance between the aortic valve and the cardiac apex) of >1000 frames/s was acquired using the ECG-gated kilohertz visualization technique. The probe was then rotated 90° and positioned 6 mm below the mitral annulus. Three parasternal short-axis (SAX) M-mode sequences were stored. Percent fractional shortening (% FS) was calculated in the M-mode image as FS = (EDD − ESD) / EDD, where EDD and ESD are the end-diastolic and end-systolic diameters, respectively.

2.3. Lung wet-to-dry weight ratio

Lung tissues were harvested, blotted dry and weighed in order to de-
termine the weight of the lung in the wet state and the wet-to-dry weight ratio was calculated, as follows: the lung tissues were weighed; wrapped loosely in aluminum foil; dried in an oven at 80 °C for 24 h; and weighed again. Then the ratio lung wet-to-dry weight ratio was calculated.

2.4. Histopathology examination

After tissue harvest, the lungs were fixed in 4% buffered formalin so-
olution, dehydrated, embedded in paraffin, and then sliced into 5-mm-thick sections to evaluate lung micro-morphological injury. After deparaffinization, tissue sections were stained using standard hematoxy-
lin and eosin (HE) staining method.

2.5. Enzyme-linked immunosorbent assay

The concentration of each respective protein/peptide, as described below under the plasma/serum and pulmonary tissue extracts section, was determined using the following kits: serum and lung levels of tumor necrosis factor alpha (TNF-α) and ET-1; plasma levels of IL-6 (R & D Systems, Minneapolis, MN), according to the manufacturer’s proto-
ocol. These kits are already validated for the measurement of rat samples in our lab.

2.6. RNA preparation and real-time quantitative polymerase chain reaction

Total RNA samples from lung tissues were isolated using the acid guanidinium thiocyanate phenol chloroform extraction with RNaseA (Qiagen, Tokyo, Japan). After RNA isolation, quantification and DNase I
treatment, RNA was reverse transcribed to cDNA using Omniscript RT and the first-strand cDNA synthesis kit (Qiagen). The reaction was performed at 37 °C for 60 min.

The mRNA expression levels of target genes were analyzed by real-time quantitative PCR using TaqMan probe and an ABI Prism 7700 sequence detector (PerkinElmer Applied Biosystems, Foster, CA). The gene-specific primers and TaqMan probes were synthesized from Primer Express version 1.5 software (PerkinElmer), according to the published cDNA sequences for each gene, as previously described [32,37]. The PCR mixture (25 μl total volume) consisted of forward and reverse primers for each gene (PerkinElmer) at 450 nM each, FAM-labeled primer probes (PerkinElmer) at 200 nM, and RTMaster Mix (PerkinElmer). Each PCR amplification was performed in triplicate as follows: 1 cycle at 95 °C for 10 min and 40 cycles at 94 °C for 15 s and 60 °C for 1 min. The quantitative values of target mRNAs were normalized by glyceraldehyde 3-phosphate dehydrogenase (GAPDH) mRNA, because GAPDH mRNA expressions were more stable among all the samples than other internal controls, such as β-actin and 18S ribosomal RNA.

Primers and probes are as follows:

<table>
<thead>
<tr>
<th>Gene Symbol</th>
<th>Forward Primer</th>
<th>Reverse Primer</th>
<th>FAM Probe</th>
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<tr>
<td>TNF-α</td>
<td>5′-CCAGGAGAAATCGCTGCTCCT-3′</td>
<td>5′-GAAAGGCTCTGCTGTACCAGGT-3′</td>
<td>5′-TCTGCTGCTGTACCAGGT-3′</td>
</tr>
<tr>
<td>ET-1</td>
<td>5′-CCACATCCAGTGCTGCTATAA-3′</td>
<td>5′-AGAGGCCGCTCTGCTGTACCAGGT-3′</td>
<td>5′-AGAGGCCGCTCTGCTGTACCAGGT-3′</td>
</tr>
<tr>
<td>ET-A</td>
<td>5′-CCAGGAGCAGTGCTGCTATAA-3′</td>
<td>5′-AGAGGCCGCTCTGCTGTACCAGGT-3′</td>
<td>5′-AGAGGCCGCTCTGCTGTACCAGGT-3′</td>
</tr>
<tr>
<td>HIF-1α</td>
<td>5′-CTAGGAGCAGTGCTGCTATAA-3′</td>
<td>5′-AGAGGCCGCTCTGCTGTACCAGGT-3′</td>
<td>5′-AGAGGCCGCTCTGCTGTACCAGGT-3′</td>
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2.7. Statistical analysis

The results were expressed as mean ± SE, and for parametric distribution, analysis of variance test was used. For non-parametric distribution, Kruskal-Wallis test, followed by Steel-Dwass for multiple comparisons was used. A p < 0.05 was considered statistically significant for all parameters.

3. Result

3.1. Blood gas analysis results

Table 1 shows the data of blood gas analysis in the current experimental setting. pH was not significantly different among the three groups (Table 1). Arterial PaO₂ was significantly decreased in LPS-administered rats compared to control group and landiolol treatment significantly increased the levels of arterial PaO₂ in septic rats (Table 1). However, arterial PaCO₂ was not significantly changed among the three groups. Blood lactate concentrations were elevated with LPS administration and 3 h treatment with landiolol partly normalized the elevated levels of lactate in LPS-administered rats (Table 1). HCO₃⁻ and base excess were not significantly changed among the three groups (Table 1).

Table 1  
<table>
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<th>Parameters</th>
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<th>LPS + landiolol</th>
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<tbody>
<tr>
<td>pH</td>
<td>7.39 ± 0.01</td>
<td>7.37 ± 0.02</td>
<td>7.38 ± 0.01</td>
</tr>
<tr>
<td>PaO₂ (Torr)</td>
<td>102.8 ± 2.2</td>
<td>84.2 ± 4.7*</td>
<td>102.4 ± 3.9*</td>
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<tr>
<td>PaCO₂ (Torr)</td>
<td>40.2 ± 1.4</td>
<td>40.9 ± 2</td>
<td>37.2 ± 2</td>
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<tr>
<td>Lactate (mmol/l)</td>
<td>0.5 ± 0.5</td>
<td>-2.8 ± 0.7</td>
<td>-3.9 ± 0.7*</td>
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<tr>
<td>HCO₃⁻ (mmol/l)</td>
<td>25.3 ± 0.5</td>
<td>21.4 ± 0.7*</td>
<td>20.7 ± 0.8*</td>
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</table>

Data are mean ± SE. Abbreviations: LPS, lipopolysaccharide.  
* p < 0.05 vs. control.  
♯ p < 0.05 vs. LPS.

3.2. Hemodynamic changes in current experiments:

We assessed the hemodynamic parameters in the rats after LPS administration. As shown in Table 2, both the levels of the systolic and diastolic blood pressure were significantly lower at 3 h after LPS administration compared to that of the control group and was unaffected by landiolol treatment. Landiolol significantly decreased the elevated heart rate in LPS-administered animals (Table 2). % FS was also significantly increased (hyperkinetic) in LPS-administered group compared to that of control group, and a 3 h long treatment of LPS-administered rats with landiolol significantly reversed the elevated % FS (Table 2).

Table 2  
<table>
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<th>Parameters</th>
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<th>LPS + landiolol</th>
</tr>
</thead>
<tbody>
<tr>
<td>Systolic BP (mm Hg)</td>
<td>125.6 ± 3.2</td>
<td>93.2 ± 6.6*</td>
<td>98.5 ± 5*</td>
</tr>
<tr>
<td>Diastolic BP (mm Hg)</td>
<td>99.8 ± 3.1</td>
<td>72.3 ± 5.8*</td>
<td>75.5 ± 4.8*</td>
</tr>
<tr>
<td>Heart rate (bpm)</td>
<td>468.7 ± 19</td>
<td>501.2 ± 16.2*</td>
<td>439.9 ± 6.5*</td>
</tr>
<tr>
<td>Fractional shortening (%)</td>
<td>40.3 ± 1.3</td>
<td>45.1 ± 1.2*</td>
<td>41.2 ± 1*</td>
</tr>
</tbody>
</table>

Data are mean ± SE. Abbreviations: BP, blood pressure; LPS, lipopolysaccharide.  
* p < 0.05 vs. control.  
♯ p < 0.05 vs. LPS.

3.3. Morphological evaluation

In the LPS-administered group, lung tissues showed thickening of alveolar, congestion formation and infiltration by neutrophils, but (these morphological abnormalities) were absent in the lung of the control group (Fig. 1A). Landiolol treatment for 3 h significantly improved the lung congestion in septic rats (Fig. 1A).

3.4. Evaluation of expression of ET-1 system in lung tissue

Levels of ET-1 peptide were also evaluated in the current experimental setting (Fig. 2A). ET-1 peptide expression was significantly increased in the lung tissue (about 2-fold) after LPS administration compared to the control group and landiolol treatment for 3 h significantly normalizing the elevated levels of pulmonary ET-1 peptide in septic rats (Fig. 2A). While levels of pulmonary ET-1 in septic rats was greatly ameliorated following the 3 h treatment by landiolol, landiolol treatment failed to normalize the elevated levels of plasma ET-1 in septic rats (Table 3). Prepro ET-1 mRNA levels were also elevated in lung tissue.
in the LPS-administered group compared to the control group. However, landiolol treatment normalized the elevated levels of prepro ET-1 mRNA in septic lung tissues (Fig. 2B). Moreover, ET-A receptor mRNA expression in lung tissue was also elevated in the LPS administration group compared to the control group, and landiolol treatment normalized the elevated levels of ET-A receptor mRNA (Fig. 2C) in septic rats. In contrast, ET-B receptor mRNA expression in lung tissue significantly decreased in the LPS administration group compared to the control group, and landiolol treatment failed to normalize the decreased levels of ET-B receptor mRNA in septic pulmonary tissues (Fig. 2D).

Fig. 1. A; Morphological findings by hematoxylin and eosin staining (HE) in lung tissues in control rats, 3 h after lipopolysaccharide (LPS) administration, and 3 h after LPS plus landiolol hydrochloride administration. Magnification, ×320. B; Pulmonary wet-to-dry weight ratio in control rats, 3 h after lipopolysaccharide (LPS) administration, and 3 h after LPS plus landiolol hydrochloride administration. Wet-to-dry weight ratios were determined for each experimental group. Values are mean ± SE (n = 5). * p < 0.05 vs. control, # p < 0.05 vs. LPS.

Fig. 2. Peptide and mRNA expression levels of ET-1 (A and B), mRNA expression levels of ET-A (C), ET-B (D) receptors in lung tissues in control, 3 h LPS-administered rats, and landiolol treated 3 h LPS-administered rats. ET-1 peptide was measured by ELIZA. mRNA expression level was determined by Real Time PCR. The control was normalized as 100%. Values are mean ± SE (n = 15). * p < 0.05 vs. control, # p < 0.05 vs. LPS.
expression levels were also increased in the LPS administration group.

Data are mean ± SE. Abbreviations: LPS; lipopolysaccharide.

3.6. Evaluation of expression levels of HIF-1α

Both the upregulated plasma levels of potential inflammatory cytokines, TNF-α and IL-6 were unchanged with the treatment of landiolol in septic rats (Table 3). We also evaluated the expression patterns of mRNA and protein of TNF-α, the key inflammatory cytokine in lung tissues during sepsis. Levels of both protein and mRNA (TNF-α) were significantly higher in septic lungs compared to the control group and a 3 h treatment of septic rats with landiolol could not normalize the elevated levels of TNF in lungs (Fig. 3A, B). In the current experimental setting, the expression levels of IL-6, another highly potent inflammatory cytokine, had parallel expression pattern as TNF-α (Fig. 3C), which was up-regulated in LPS-administered rats and was unchanged following landiolol treatment.

3.5. Evaluation of expression levels of inflammatory cytokine (TNF-alpha and IL-6)

In addition, to clarify whether LPS administration induces any hypoxic condition in pulmonary tissues, we examined the mRNA expression levels of HIF-1α in lung tissue in present study. HIF-1α mRNA expression levels were also increased in the LPS administration group in lung tissues, but landiolol treatment did not normalize the elevated pulmonary levels of HIF-1α (Fig. 3D).

4. Discussion

The present study is the first to demonstrate the normalization of elevated ET levels in rat pulmonary tissues by an ultrashort acting beta blocker (landiolol hydrochloride) during the early stages (hours) of sepsis. Further, this phenomenon (of landiolol on pulmonary ET-1 level) is also accompanied by an improvement in the morphology of the septic lung tissues. Here, we also show that pre-treatment of LPS-administered rats with landiolol for 3 h normalized both the diminished levels of PaO2 and enhanced levels of blood lactate in early sepsis. In contrast, these observed pulmonary ameliorations (molecular and morphological) in septic rats induced by landiolol are not associated with alteration in levels of inflammatory cytokines (TNF-α, IL-6) and hypoxia marker (HIF-1α) under the current experimental setting.

A previous study has reported that LPS increases heart rate and %FS in rats 3 h post administration and that if these (septic) animals are treated with landiolol these altered conditions (HR and %FS) are normalized [28,29]. It is interesting that while landiolol was able to decrease %FS and HR in septic rats, as reported previously, it (landiolol) had no significant effect on systolic and diastolic blood pressure under the current experimental setting. In fact, landiolol’s effect on blood pressure was lesser than esmolol, an ultra-short-acting β-blocker [38,39]. It is likely that landiolol’s ability to attenuate the LPS-induced hyperdynamic state in septic rats may be linked to its (landiolol) partial normalization of elevated blood lactate concentrations in early sepsis.

The present study demonstrates landiolol’s ability to improve PaO2 during the early phase (hours) of sepsis in rats, as well as reduction of congestion and normalization of the wet-to-dry ratio in lungs compared to the untreated group. These findings are significant since LPS has been shown to induce edema formation, infiltration of interstitial tissue by

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Table 3

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<thead>
<tr>
<th></th>
<th>Control</th>
<th>LPS</th>
<th>LPS + landiolol</th>
</tr>
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<tbody>
<tr>
<td>TNF-α (plasma, pg/ml)</td>
<td>15.7 ± 7**</td>
<td>180 ± 40**</td>
<td>184.3 ± 30.1**</td>
</tr>
<tr>
<td>ET-1 (plasma, pg/ml)</td>
<td>3.8 ± 0.4*</td>
<td>34 ± 3.3*</td>
<td>32.5 ± 3.3*</td>
</tr>
<tr>
<td>IL-6 (plasma, pg/ml)</td>
<td>18.3 ± 1.9*</td>
<td>29 ± 3.3*</td>
<td>26.7 ± 3.7*</td>
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</table>

Data are mean ± SE. Abbreviations: LPS; lipopolysaccharide.

* p < 0.05 vs. control.
** p < 0.05 vs. LPS.

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Fig. 3. mRNA and protein expression levels of TNF-α (A and B), and mRNA expression levels of IL-6 (C), HIF-1α (D) in lung tissues in control, 3 h LPS-administered rats, and landiolol treated 3 h LPS-administered rats. TNF-α protein was measured by ELIZA. mRNA expression level was determined by Real Time PCR. The control was normalized as 100%. Values are mean ± SE (n = 15). * p < 0.05 vs. control, # p < 0.05 vs. LPS.
neutrophils, and reduction of alveolar spaces in lungs, 12 h post-treatment [40] and, importantly, these (same) morphological injuries diminished after administration of landiolol [26]. Thus, based on the current data, we speculate that landiolol is equally effective in eliminating acute septic-induced pulmonary tissue injuries through the reduction of congestion, and, ultimately, leads to a decrease in pulmonary wet-to-dry weight ratio and improvement in arterial PaO₂.

ET-1 is a potent vasoconstrictor and pro-inflammatory peptide and has previously been implicated in the pathogenesis of sepsis and sepsis-induced MODS [41,42]. Therefore, the present data, showing an increase in the plasma levels of ET-1, are consistent with data obtained from earlier studies [35] from our lab and others [16,17,43]. This observed increase in levels of plasma ET-1 in septic animals may be due to a combination of enhanced ET-1 synthesis, as well as impairment in ET clearance in renal and pulmonary tissues [16]. ETs is also known to serve increase in levels of plasma ET-1 in septic animals may be due to a combination of enhanced ET-1 synthesis, as well as impairment in ET clearance in renal and pulmonary tissues [16]. ETs is also known to serve increase in levels of plasma ET-1 in septic animals may be due to a combination of enhanced ET-1 synthesis, as well as impairment in ET clearance in renal and pulmonary tissues [16]. 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